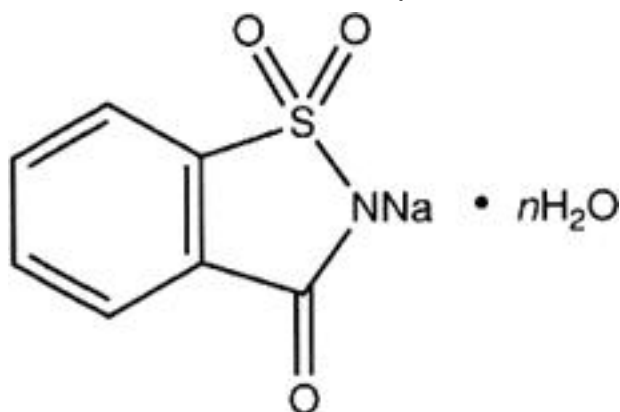


**Saccharin sodium (Saccharinum natricum)****Saccharin sodium, anhydrous****Saccharin sodium, dihydrate** $n = 0$  (anhydrous) $n = 2$  (dihydrate) $\text{C}_7\text{H}_4\text{NNaO}_3\text{S}$  (anhydrous) $\text{C}_7\text{H}_4\text{NNaO}_3\text{S} \cdot 2\text{H}_2\text{O}$  (dihydrate)**Relative molecular mass.** 205.2 (anhydrous); 241.2 (dihydrate).**Chemical name.** 1,2-Benzisothiazolin-3-one 1,1-dioxide, sodium salt; 1,2-benzisothiazol-3(2H)-one 1,1-dioxide, sodium salt; CAS Reg. No. 128-44-9 (anhydrous).

1,2-Benzisothiazolin-3-one 1,1-dioxide, sodium salt, dihydrate; 1,2-benzisothiazol-3(2H)-one 1,1-dioxide, sodium salt, dihydrate; CAS Reg. No. 6155-57-3 (dihydrate).

**Other name.** Saccharimidum natricum.**Description.** Colourless crystals or a white, crystalline powder; odourless or almost odourless.**Solubility.** Freely soluble in water; sparingly soluble in ethanol (~750 g/l) TS; practically insoluble in ether R.**Category.** Sweetening agent.**Storage.** Saccharin sodium should be kept in a well-closed container.**Additional information.** Saccharin sodium effloresces slowly in air and loses about half of its content of water of crystallization. It has a very sweet taste, even in very dilute solutions.**Requirements**Saccharin sodium contains not less than **98.0%** and not more than the equivalent of **101.0%** of  $\text{C}_7\text{H}_4\text{NNaO}_3\text{S}$ , calculated with reference to the anhydrous substance.**Identity tests**

A. To 20 mg add 0.04 g of resorcinol R and 0.5 mL of sulfuric acid (~1760 g/l) TS, and heat gently until a dark green colour is observed. Allow to cool and add 10 mL of water and 10 mL of sodium hydroxide (~80 g/l) TS; a fluorescent green solution is produced.

B. Ignite 1 g and proceed with the residue as follows:

- Dissolve half of the residue in acetic acid (~60 g/l) TS. When tested for sodium as described under [2.1 General identification tests](#) it yields reaction B.
- Dissolve the remaining residue in hydrochloric acid (~70 g/l) TS. It yields reaction A described under [2.1 General identification tests](#) as characteristic of sulfates.

**Arsenic.** Transfer 3.3 g to a crucible containing 3.3 g of anhydrous sodium carbonate R. Moisten with a small quantity of water, evaporate to dryness on a water-bath, and ignite to 550 °C until all black particles have disappeared. Cool, dissolve the residue in 5 mL of hydrochloric acid (~250 g/l) AsTS, and proceed as described under [2.2.5 Limit test for arsenic](#); the arsenic content is not

more than 3 µg/g.

**Heavy metals.** Use 1.0 g for the preparation of the test solution as described under [2.2.3 Limit test for heavy metals](#), Procedure 1; determine the heavy metals content according to Method A; not more than 20 µg/g.

**Water.** Determine as described under [2.8 Determination of water by the Karl Fischer method](#), Method A, using 1 g of Saccharin sodium dihydrate; the water content is not more than 150 mg/g.

**Free acid or alkali.** Dissolve 1 g in 10 mL of carbon-dioxide-free water R, add 5 mL of sulfuric acid (0.005 mol/l) VS, boil, cool, and titrate with sodium hydroxide (0.01 mol/l) VS using phenolphthalein/ethanol TS as indicator; 4.5-5.5 mL are required to obtain a pink colour.

**Related substances.** Carry out the test as described under [1.14.1 Chromatography, Thin-layer chromatography](#), using silica gel R1 as the coating substance and a mixture of 100 volumes of chloroform R, 50 volumes of methanol R, and 10 volumes of ammonia (~260 g/l) TS as the mobile phase. Apply separately to the plate 5 µl of each of the following four solutions: For solution (A) dissolve 2.6 g of Saccharin sodium in 10 mL of sodium hydrogen carbonate (100 g/l) TS, add 12.5 g of diatomaceous support R as a filter-aid, and mix well. Transfer to a chromatographic tube, 250 mm in length and with a diameter of 25 mm, fitted at the lower end with a sintered-glass disc and a stopcock. Pack the contents of the tube by tapping on a padded surface and tamping firmly from the top. Elute with dichloromethane R at a rate of 50 mL in 30 minutes. Evaporate the eluate to dryness and dissolve the residue in 4 mL of acetone R. For solutions (B) dissolve 50 µg of toluene-2-sulfonamide RS per mL of acetone R, (C) 5 mg of Saccharin sodium per mL of methanol R, and (D) 50 µg of 4-sulfamoylbenzoic acid R per mL of acetone R. After removing the plate from the chromatographic chamber, dry in a current of warm air, heat at 105 °C for 5 minutes, and spray the hot plate with sodium hypochlorite TS1. Dry in a current of cold air until a sprayed area of the plate below the line of application gives at most a faint blue colour with 0.05 mL of a solution of 5 mg of potassium iodide R in 1 mL of starch TS containing 1% glacial acetic acid R. Avoid prolonged exposure to cold air. Spray the plate again with the same mixture. Examine the chromatogram in daylight.

Any spot obtained with solution A corresponding to toluene-2-sulfonamide is not more intense than that obtained with solution B. Any spot obtained with solution C corresponding to 4-sulfamoylbenzoic acid is not more intense than that obtained with solution D.

**Assay.** Dissolve about 0.3 g, accurately weighed, in 30 mL of glacial acetic acid R1, and titrate with perchloric acid (0.1 mol/l) VS as described under [2.6 Non-aqueous titration](#), Method A.

Each mL of perchloric acid (0.1 mol/l) VS is equivalent to 20.52 mg of C<sub>7</sub>H<sub>4</sub>NNaO<sub>3</sub>S.